

Intraoperative neuromonitoring techniques in the surgical management of acoustic neuromas

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Unfavorable outcomes such as facial paralysis and deafness were once unfortunate probable complications following resection of acoustic neuromas. However, the implementation of intraoperative neuromonitoring during acoustic neuroma surgery has demonstrated placing more emphasis on quality of life and preserving neurological function. A modern review demonstrates a great degree of recent success in this regard. In facial nerve monitoring, the use of modern electromyography along with improvements in microneurosurgery has significantly improved preservation. Recent studies have evaluated the use of video monitoring as an adjunctive tool to further improve outcomes for patients undergoing surgery. Vestibulocochlear nerve monitoring has also been extensively studied, with the most popular techniques including brainstem auditory evoked potential monitoring, electrocochleography, and direct compound nerve action potential monitoring. Among them, direct recording remains the most promising and preferred monitoring method for functional acoustic preservation. However, when compared with postoperative facial nerve function, the hearing preservation is only maintained at a lower rate. Here, the authors analyze the major intraoperative neuromonitoring techniques available for acoustic neuroma resection.
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hearing preservation • facial nerve preservation

ACOUSTIC neuromas (vestibular schwannomas) are categorized as benign, extraaxial brain tumors (Fig. 1) developing near the internal auditory canal, typically with involvement of the cerebellopontine angle.^{32,53,60,130,143} Advances in treatment modalities have popularized the application of less invasive management methods such as radiotherapy and radiosurgery,^{100,138} which carry high efficacy and low morbidity.^{31,53,57,86,87,100,101,107} However, many acoustic neuromas, particularly those that are large in size, necessitate surgical intervention.^{33,36,62,101,107,110}

The primary operative goals are gross tumor debulking while safeguarding the adjacent cranial nerves (Fig. 1).^{4,9,36,49,103,115,118,124,130} Neural preservation is particularly im-

perative in the contemporary management of acoustic neuromas.^{5,68,103} By virtue of their location, these tumors are close to the facial and vestibulocochlear cranial nerves (Fig. 1), and can thus severely impair the nerve function at the time of initial presentation.^{60,62,68,79,143} The neuroma can directly impinge, tightly adhere to, or overtly damage the nerves.^{13,22,38,60} These tumors often present as operative challenges, as resection may cause nerve irritation or injury leading to neurapraxia, axonotmesis, or neurotmesis.^{38,117}

The various options of surgical approaches (translabyrinthine vs middle fossa vs retrosigmoid) for acoustic neuromas and their respective patterns of postoperative cranial nerve preservation have been described.^{3-6,15,23,25,45,49,56,58,61,65,67,112,115,117,118,124,130,131} However, IONM may demonstrate improvements in structural and functional preservation of the cranial nerves during these operations.^{139,148} Several IONM techniques have been developed and evaluated with particular focus on CN VII and VIII preservation. Among these methods, the most frequently

Abbreviations used in this paper: BAEP = brainstem auditory evoked potential; CN = cranial nerve; CNAP = compound nerve action potential; ECOG = electrocochleography; EMG = electromyography; IONM = intraoperative neuromonitoring; IOVM = intraoperative video monitoring; MUP = motor unit potential.

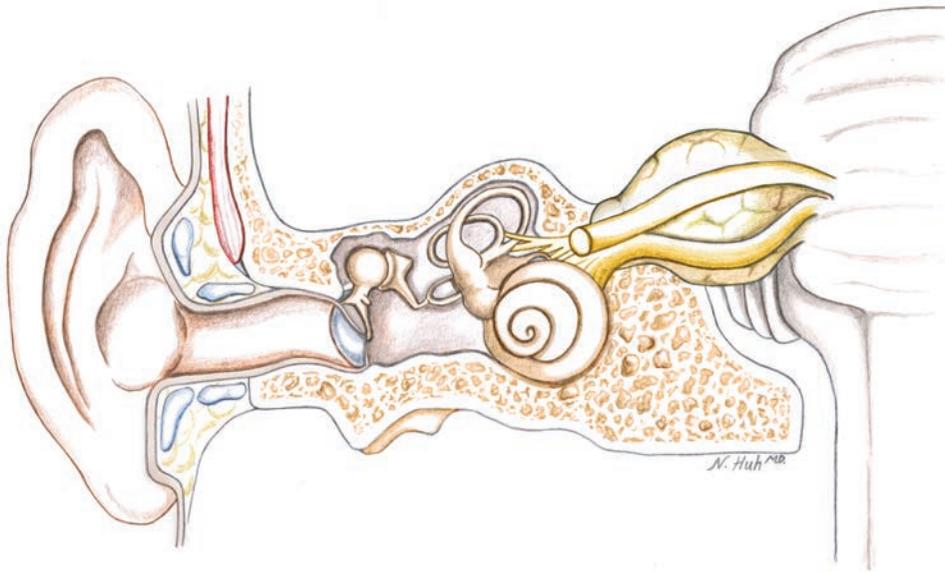


Fig. 1. Illustration showing an acoustic neuroma with displaced facial and cochlear nerves, the nerves we are trying to preserve. Printed with permission from Dr. Nancy Huh, M.D., Illustrations.

used are EMG for the facial nerve and BAEP monitoring for the vestibulocochlear nerve.^{41,42,79,142,146,148} Here, we assess the fundamental characteristics underlying the major techniques available in IONM, emphasizing specific advantages and limitations of their utilization for optimal patient management.

Intraoperative Monitoring of the Facial Nerve (CN VII)

Cranial nerve VII plays a critical role in facial muscle function and one's cosmetic appearance, and its weakness can have severe and profound implications on a patient's quality of life.^{72,92,140,148} For instance, loss of facial nerve function can ultimately result in an inability to blink, secrete tears, or speak properly, thus imposing a significant burden on the patient.^{6,85} Such significant outcomes were once considered a probable morbidity.^{78,92,117} However, with the advent of facial neuromonitoring, the morbidity once associated with acoustic neuroma resection has been drastically reduced. The House-Brackmann Grading Scale,⁴³ which ranges in increasing severity of deficits from Grade I through Grade VI, serves as a standardized method for analyzing postoperative outcomes of facial nerve function. As a result of advances in microsurgery and facial nerve IONM, many patients with smaller tumors have minimal functional loss of the nerve, as indicated by low House-Brackmann grades.^{3-7,12,27,36,46,62,110,115,117,124,130,146} In patients with larger tumors, the outcomes are not as optimistic, as these patients are at an increased risk of postoperative facial nerve deficits.^{4,27,36,56,60,62,117,125}

Electromyography

The use of EMG to monitor facial nerve function has been well documented, leading to its widespread application in modern practice.^{9,22,39,40,46,54,82,94,125,126} The operative

EMG device consists of a stimulator probe and a "sensor" that detects contractions of the facial muscles. Most operations use a minimum of 2 channels to observe the activity of the orbicularis oris and orbicularis oculi muscles,^{12,46,77,82,90,115} although the use of additional channels to observe other facial muscles may provide further benefit.^{37,38,85,134} When considering a 2-channel system, a pair of needle electrodes are usually planted in the orbicularis oris and orbicularis oculi muscles while another is placed on the forehead or shoulder for grounding.^{9,12,54,82} Prior to the operation, the baseline electrical parameters, including MUPs and insertional activity, of these muscles are measured and recorded for future comparisons.^{22,78}

The stimulator probe is applied to determine the location of the facial nerve.^{21,40,126,148} During an operation, the ideal location for applying the probe on the facial nerve is near the brainstem^{13,37} because it is proximal to the area of resection. Distal stimulation, while possible, yields limited data, as stimulation is being directed on the portion of the nerve that is virtually unaffected by resection.³⁵ However, distal stimulation is not to be ignored, as several studies have found that higher proximal-to-distal EMG amplitude ratios successfully predict postoperative facial nerve function.^{35,46,47,137} When delivering the stimulus, the amount of current that is administered by the probe can be adjusted.^{21,22,38,70,90,126} Once the amount of current applied exceeds the action potential threshold of the patient's facial nerve,¹¹¹ an action potential is fired that causes twitching of the facial muscles.^{22,126} The sensor detects these facial movements and emits a sound alarm, thereby providing direct, immediate, and real-time feedback.^{9,34,66,111,126} The facial muscle MUPs corresponding to this stimulation are also projected onto an oscilloscope to facilitate visualization.^{39,42,82}

The electrical morphology, frequency, and characteristics of the MUPs vary greatly, and such divergences offer insights into possible abnormal nerve activity.^{42,77} As demonstrated in Fig. 2,¹¹¹ multiple types of MUP sig-

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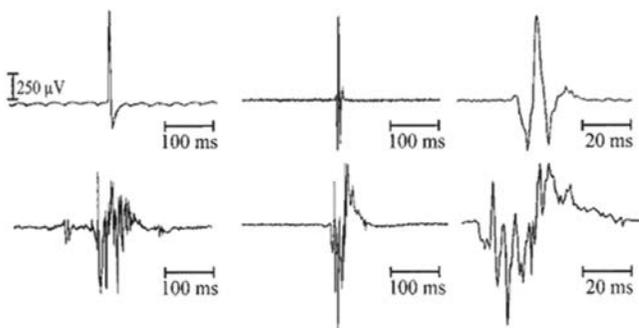


Fig. 2. Electromyography MUP morphologies demonstrating spikes (*upper*) and bursts (*lower*). Reprinted with permission from Romstöck et al: *J Neurosurg* 93:586–593, 2000.

nals can be observed on an intraoperative EMG study. A single MUP wave is referred to as a “spike,” while a short chain of MUPs is classified as a “burst.” When a sustained streak of MUPs is distinguished, it is designated as a “train,”⁷⁷ which is shown in Fig. 3.¹¹¹ Train MUPs possessing a particularly high frequency (greater than 30 Hz) are termed “neurotonic.”^{74,1,42,77}

Neurotonic train activity typically serves as an indicator of intense nerve stimulation, as robust nerve stimulation correlates with greater MUP activation.⁷⁷ During an operation, neurotonic discharges can occur in the context of nerve stimulation, irritation, or damage.^{41,42,148} However, not all neurotonic train waves carry equal clinical significance. The A-train pattern has been most substantially affiliated with postoperative facial nerve deficits (Fig. 3).¹¹¹ The A trains are characterized as a high-frequency train pattern with the following features: a duration lasting milliseconds to seconds, an amplitude in the range of 100–200 μ V, and a short onset and offset.^{104,105,111,148} The duration of “train time,” as quantified by the seconds of A-train activity, has been shown to translate to worse postoperative facial nerve paresis.^{21,104,105} Other train patterns that may be encountered on an EMG include the B and C trains, although they have not been shown to carry significant value

in predicting postoperative nerve function.¹¹¹ As described by Romstöck et al.,¹¹¹ B trains manifest either in a spike or burst pattern and are distinguished by their gradual onset, low amplitudes, and average duration lasting minutes to hours. C trains, on the other hand, are irregular waveforms of varying amplitudes that bear resemblance to interference. Aside from train time and activity, other electrical EMG findings bear clinical importance as well. Mandpe et al.⁶⁶ reported that low immediate postoperative stimulation thresholds in combination with a response amplitude appeared to reliably foretell excellent postoperative facial nerve function. Neff et al.⁹⁰ reached similar conclusions but with stimulation thresholds of 0.05 mA or lower and amplitudes greater than 240 μ V.

Electromyography provides several benefits. One of its main functions is determining the anatomical location of the facial nerve.^{22,126} Direct, pinpoint visualization of the nerve may often prove difficult, as the tumor, its capsule, and bone may interject along the nerve’s trajectory. By adjusting and determining the current required for muscle stimulation, however, the relative proximity of the nerve to the probe can be deduced. If there is very little tumor, tissue, or bone covering the nerve, the facial nerve will be more prone to stimulation at lower currents, such as lower than 0.2 mA, thereby implying that the nerve is highly exposed, close to the probe, and in danger of being manipulated.^{22,126} Conversely, stimulation at higher currents, such as greater than 0.5 mA, suggests the presence of a sizable tissue or bone barrier between the nerve from the probe.¹²⁶ Highly adherent tumors have a tendency for creating thicker barriers from the probe, thus resulting in higher mean stimulation thresholds.³⁸

In addition, EMG helps prevent unplanned manipulation of the facial nerve by emitting a warning noise whenever muscle stimulation is detected. This can warn the surgeon of impending danger and thus advise cessation of current actions or recommend extreme caution. By doing so, EMG directly influences surgical planning and strategy, as the surgeon can appropriately alter the surgical ap-

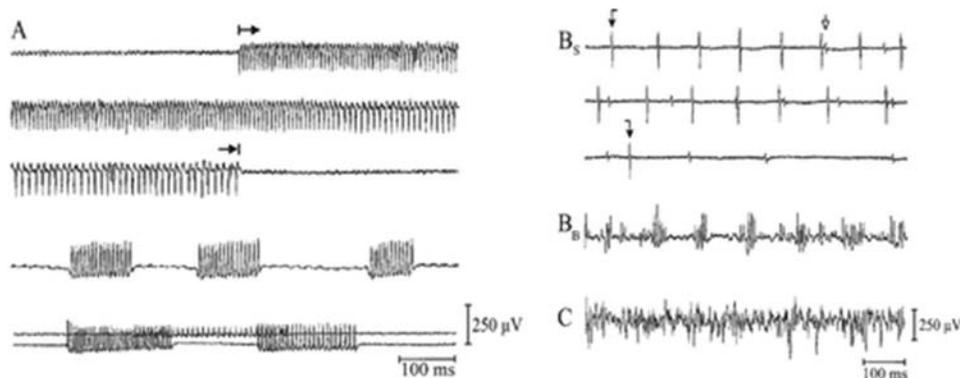


Fig. 3. Electromyographic train activity. **Left:** Examples of A trains of various durations and frequencies. The *upper* tracings show the abrupt onset and termination (*arrows*) of this sinusoidal waveform pattern, which lasted 1600 msec. The fourth train from the top of the figure shows repeated short-term periods of activity, ranging in duration from 100 to 120 msec each. The *lower* A train, which was simultaneously recorded from 2 facial muscle groups, gives an impression of frequency variability between 120 and 190 Hz. **Right:** Waveforms defined as B trains with spikes (B_s) and B trains with bursts (B_b) as predominant single components. The *black* and *white* arrows mark 2 individual B trains with spikes of higher and lower amplitudes recorded in the same channel. The *lowest* tracing represents irregular EMG activity, called a C train. Reprinted with permission from Romstöck et al: *J Neurosurg* 93:586–593, 2000.

proach to avoid causing damage to the nerve.^{9,25,46,82,111,125} A-train activity or other abnormal EMG patterns may also encourage caution,³⁹ although they must be placed in context: neurotonic discharges can sometimes fire even in the presence of a transected nerve.⁴² In addition to perioperative nerve preservation, EMG can help clarify the residual function of the nerve postoperatively.^{46,54} When comparing postoperative and baseline stimulation thresholds, patients who require high or higher postoperative currents may have endured some degree of nerve injury.^{54,70,93,123,126,128,150}

Despite its benefits, EMG is not an infallible monitoring system. During resection, the facial nerve may appear grossly intact; however, this finding does not necessarily convert to true nerve functionality.^{11,22,60,70,92} One possible explanation for this phenomenon is that EMG can sometimes receive poor data input. This issue is particularly salient with the application of microinstruments to cauterize tissue or tumor surrounding the facial nerve.^{41,85} The generated electrical signal may create artifact, signal interference, and distortion.

Electromyography also runs the added risk of instigating electrical injury from overstimulation. As general principle, application of the stimulator probe should be done conservatively to avoid inducing iatrogenic injury to the facial nerve. Intense or prolonged stimulation theoretically increases the risk of causing irreparable nerve injury.^{106,127} To that end, several techniques are encouraged to diminish the risk of injury. Pulsed stimulation, for example, appears to have a lower injury risk than constant stimulation.^{26,106} In addition, monopolar stimulation with constant voltage may be superior to bipolar constant-current stimulation.⁸² However, the majority of experimental studies done to examine the potential for overstimulation have been conducted in animal models. Through these studies, one of the emerging general principles has been the greater influence of stimulus frequency on the degree of nerve injury. In rats, Sapmaz et al.¹¹⁹ investigated the respective effects of stimulus amplitude (mA) and frequency on histological axonal degeneration. The authors' results demonstrated that frequency, but not amplitude, was statistically significant in causing greater axonal degeneration. In other words, rats with 20 stimulations had more degeneration than those that underwent 10 stimulations ($p < 0.05$), while rats with 30 stimulations had more degeneration than those that received 20 stimulations ($p < 0.05$).¹¹⁹ In a cat model, McCreery et al.⁷⁶ obtained similar results: stimulation at 100 Hz versus 50 Hz caused greater axonal degeneration, while stimulus amplitude did not appear to have much effect. Another important principle is the superiority of pulsed stimulation when compared with constant stimulation. In mice, pulsed stimulation was associated with less myelin and axonal degeneration.⁴⁴ In a cat study, investigators found that extended periods of high frequency stimulation caused greater injury and that pulsed stimulations can reduce the risk of damage.² Interestingly, Kartush et al.⁵⁵ found that constant current stimulation can be safely applied in guinea pig models as long as the electrode is properly insulated to preclude shunting. In summary, low stimulus frequencies and pulsed stimulations can be applied clinically to minimize the risk of injury from overstimulation.

Direct Observation/Video Monitoring

To increase the sensitivity of facial nerve IONM, recent studies have proposed implementing direct observation of facial muscle movement or intraoperative video monitoring (IOVM).^{21,28,29,85} Theoretically, IOVM would supplement EMG by allowing better visualization of facial muscle contractions, thus providing an additional aid in the operating room. During IOVM, an anesthesia mask containing several infrared cameras is fastened to the patient's face, and the infrared properties of these cameras allow video recording under the operative drapes.^{28,85} The camera view can be magnified such that even minute movements may be detected by the naked eye.⁸⁵ The images are projected on a 4-way split screen: 2 focus on movements of the facial muscles, another displays the microscopic operating field, and the remaining screen projects the EMG tracings.^{28,29} These simultaneously derived images are thus juxtaposed next to each other, with a sound alarm triggered by facial muscle contractions.^{28,29}

Although IOVM may prove useful, the full utility of this tool remains to be characterized. In a study comparing EMG with IOVM, the use of EMG alone exhibited higher sensitivity in detecting facial nerve activation: EMG detected facial muscle movement at a stimulation of 0.3 mA, whereas IOVM required a minimum of 0.5 mA.²⁹ De Seta et al.²¹ obtained similar results, finding EMG alone to be more sensitive than IOVM. Thus, EMG appears more effective than IOVM based on current data. However, further studies must evaluate the validity of IOVM as a supplementary tool in the operating room.

Intraoperative Monitoring of the Vestibulocochlear Nerve (CN VIII)

Even with modern IONM, current vestibulocochlear nerve retention rates do not compare favorably with the excellent outcomes seen with the facial nerve.^{5,12,16,36,49,52,62,75,88,94,99,103,112,115,116,120,130,132,139,147} Although this discrepancy may highlight the need for improvement in IONM of CN VIII,⁹⁹ it may also reflect the inherent difficulty in preserving auditory function, as large tumors are more highly associated with postoperative deficits,^{1,11,20,23,88,89,103,116} and tumors with extensive infiltration into the cerebellopontine angle render acoustic preservation an arduous task.^{23,89,147}

Operative damage to the vestibulocochlear nerve can be induced in various ways.^{63,64,149} Direct operative trauma is a potential avenue, with the nerve most prone to exposure during maneuvers, such as drilling into the internal auditory canal, operative traction, or subsequent tumor resection.^{1,17,19,51,79,96,129} Cranial nerves are inherently more susceptible to trauma because they are ensheathed in central myelin, thus lacking the extra protective layers, such as the perineurium, that are more prevalent in peripheral myelin.^{8,17,19,64,79,123} Ischemic damage also presents further risk of injury. More specifically, vascular changes to the internal auditory artery, such as occlusion, rupture, or vasospasm, are believed to induce postoperative hearing deficits.^{17,19,79,84,88,89,122} Strauss et al.¹³⁵ found that applying medical therapy to preclude such vasospasms produced

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preservation rates that were more than twice as high when compared with controls.

Brainstem Auditory Evoked Potentials

Brainstem auditory evoked potentials are defined as the bioelectric neural activity that materializes in response to stimulation of the vestibulocochlear nerve.^{63,97,129} In comparison with the background electrical brain activity,^{54,79,142} these BAEP waves are diminutive and difficult to detect.^{18,149} To facilitate distinction between BAEPs and background "noise," several thousand samples of the electrical stimulus must be acquired and subsequently averaged to create a distinct auditory evoked potential.^{1,50,64,79,97,129,149} On BAEP recordings, the auditory response is extracted from several locations in the entire vestibular nerve pathway, as it travels peripherally to centrally.⁹⁷ The peaks of the evoked electrical potentials are classified as Wave I through Wave V, which correspond to the peripheral cochlear nerve and the inferior colliculus, respectively.^{42,52,63,64,71,97,98,129} These waves can be seen in Fig. 4.⁸⁰

In BAEP monitoring, scalp and earlobe electrodes are placed, and an auditory stimulator discharges acoustic clicks to the operated ear through an earphone-transducer apparatus.^{59,64,79,97,129,149} The electrical pulse rate is set at a range of 20–50 clicks per second.^{16,54,79,97,99,148} Before commencing with the operation, the stimulus intensity,

as measured in decibels, is adjusted until the patient can hear the click; the stimulus is eventually delivered at several decibels higher than the measured threshold.^{71,97} Upon delivery of the stimulus, the ears are stimulated bilaterally so white noise is applied at an intensity several decibels lower to obscure the response of the contralateral ear.^{50,54,64,71,96,97,121,129}

When considering BAEP waveform shifts, Waves I, III, and V carry the most clinical significance.^{54,71,74,97,129} Changes in their amplitude, peak latency, or presence of the peak are heavily scrutinized and compared with baseline BAEPs.^{23,50,54,63,71,97,98,121,129,141} More specifically, increased peak latencies of Waves I, III, and V,^{10,23,71,97,102} high interaural latency differences,^{10,23,71,97} decreased amplitudes of Waves I and V,^{63,71,97,121} and increased interpeak latencies between Waves I–III, III–V, and I–V^{71,97} are examples of potentially concerning wave changes. Between peak latencies and interpeak latencies, the latter is the more clinically useful marker because peak latencies are more susceptible to influence from external factors such as age, thus rendering them less reliable.⁷¹ However, the majority of these parameters are, at best, warning signs that alert the surgeon; among them, only maintenance of Waves I and V has been consistently shown to correlate with better postoperative hearing preservation rates,^{34,50,88,91,98,121,136,141,146} although others have found poor hearing outcomes despite wave preservation.^{30,59} The prognostic power of BAEPs is based solely on the preservation of the waves; in other words, when actual changes are seen on BAEPs, the severity or presence of postoperative deficits cannot be predicted reliably.^{18,96} Regardless, detecting such BAEP waveform irregularities can still alert the surgeon to potential cranial nerve damage and encourage redirection of the operative plan of action.⁶³

The use of BAEPs comes with several limitations. Because the stimulus response must be summed and averaged to obtain a wave of sufficiently high amplitude, the tradeoff to this process is a significant time delay that can last up to several seconds to minutes.^{1,18,19,49,50,83} Naturally, such a delay can negatively influence the course of surgery, as BAEPs effectively provide data that were applicable several seconds or minutes prior.^{14,18,108} Matthies and Samii⁷³ reported that direct BAEP monitoring was able to reduce the lag time to 5–15 seconds, suggesting that considerable improvements may be possible. In addition, BAEP recordings are prone to presenting false-positive results. Trauma is not the only causative agent of BAEP waveform shifts, with other physiological or intraoperative processes such as anesthesia, hypothermia, and irrigation all capable of inducing waveform changes.^{24,54,63,64,69,71,129,133} Such a wide range of artifact sources can create great difficulty with respect to surgical decision making.¹⁴⁵ The utility of BAEPs may also be patient dependent, as some do not have detectable BAEPs while others have abnormal baseline BAEPs.^{18,48,71,132,141} Without a clear starting point, BAEP monitoring may prove too difficult a task to complete. Measuring CNAPs may be more beneficial in such cases, as patients occasionally have BAEP waveform normalization postoperatively despite preoperative absence.¹⁰⁹

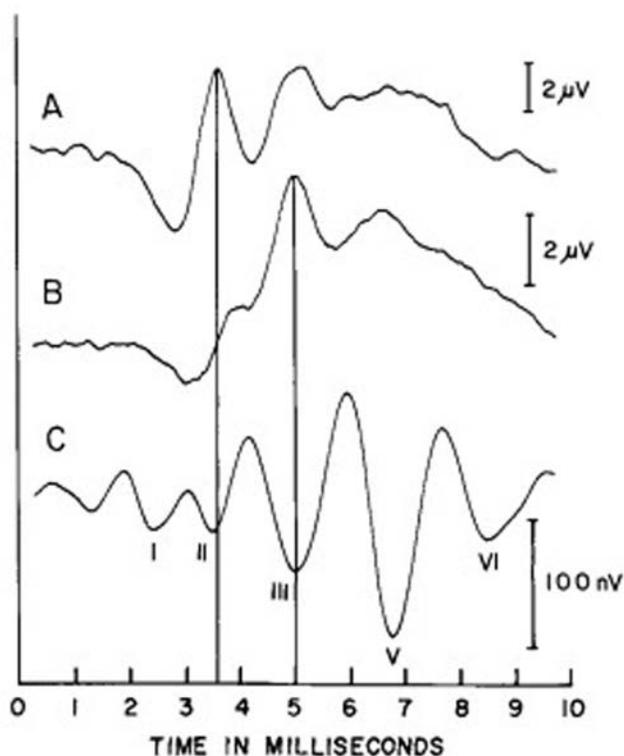


Fig. 4. Figure demonstrating direct CNAPs (A), direct recording from the lateral recess of the fourth ventricle (B), and BAEPs (C). Note the 2 negative peaks seen on direct CNAPs and the relative coincidence of the first negative peak on direct CNAP to Wave I on BAEP monitoring. Roman numerals indicate the waves. Reproduced with permission from Møller and Jannetta: *J Neurosurg* 59:1013–1018, 1983.

Electrocochleography and Direct CNAPs

Brain auditory evoked potential monitoring is considered a “far-field” technique because the auditory response is measured on the scalp, which is distal from the neural auditory response.^{54,98,129,149} In contrast, ECOG and direct CNAPs are “near-field” techniques because the stimulation evokes and records an electrical response close to its origin on the auditory nerve.^{48,149} Because these techniques record from the nerve itself, near-field IONM bypasses the noise and artifact created in far-field IONM, which translates to reducing the number of stimuli averages required in addition to affording a larger amplitude for facile visualization.^{17,54,79,149} Ultimately, this leads to a much quicker assessment of nerve function.^{114,142,149}

In principle, both ECOG and direct CNAPs use electrodes to measure potentials generated from the auditory nerve, with some minor differences in operative setup. For ECOG, electrodes are typically positioned transtympanically on the middle ear promontory of the pathological ear.^{45,53,65,80,95,96,101,123} Reference and ground electrodes are placed on the ipsilateral earlobe and on the forehead, respectively.^{64,113,114,149} A foam ear plug not only holds the electrode firmly in place but also impedes foreign substances from breaching into the ear canal.^{42,99} Similar to the BAEP, the stimulating electrode administers click impulses, and multiple responses must be averaged for a distinct wave pattern to emerge.^{18,19,113,114,120,129}

As its name suggests, in direct CNAPs, the action potential is measured directly from the acoustic nerve itself.^{64,81,129} The recording electrode is placed directly on the acoustic nerve, the negative electrode is attached to the mastoid of the contralateral ear, and a reference electrode is placed on the scalp.^{16,54,79,99,108,148} It is common practice to place the recording electrode proximal to the tumor being resected,^{14,17,79,99,129} with adhesive such as Gelfoam applied between the electrode and nerve to reinforce the placement.^{54,99,148} Like the BAEP and ECOG, a click stimulus is applied through an earphone, and the resulting compound action potential is measured.^{18,48,79,99,108,145}

Electrocochleography and direct CNAP monitoring are both techniques that rely on deducing the compound action potential, which represents a summation of all the action potentials, from the vestibulocochlear nerve.^{42,108,113,129} These CNAPs, as they are known, are visualized as negative peaks distinguishable by their high amplitudes (Fig. 4).^{80,120,129} In ECOGs, they consist of 2 action potential peaks designated “N1” and “N2,”^{113,120,142,149} and in direct CNAP monitoring, comparable peaks are obtained.^{81,108,142,149} Because ECOG involves peripheral nerve stimulation, the N1 waveform seen on ECOG is congruent with Wave I on BAEP monitoring.^{8,42,64,71,79,113,120,129} The absolute loss of N1 on ECOG^{48,88,96,114,120,139,149} or on direct recording^{8,108,129,132,145,146,149} is frequently associated with postoperative hearing deficiency. Changes to the latency or amplitude of N1 on either ECOG or direct recording are also electrophysiological signs suggestive of injury.^{8,14,17,19,48,54,79,120,131,142,145,146} Further electrical waveforms are seen in ECOG, thus differentiating it from a direct CNAP reading. The cochlear microphonics and summation potential are both electrical responses generated from the organ of Corti,^{54,94,113,120} and lower cochlear

microphonic detection thresholds may be involved in prognosticating postoperative hearing function.⁹⁴ In the overall context of ECOG monitoring, however, cochlear microphonics and summation potentials are generally considered less important than the N1 peak.^{64,120}

The primary advantages of ECOG and direct CNAPs are derived from their near-field designation. With shorter latency periods, they reflect pertinent information much faster than BAEPs and provide immediate feedback on the state of the auditory system.^{14,48,64,108,129,142,149} Changes seen on compound action potentials also tend to occur immediately, a helpful trait when considering vascular etiologies of dysfunction: vascular changes cause immediate effects that may not be detected quickly enough on BAEP monitoring.⁵⁴ The quick response time in conjunction with larger amplitudes than BAEPs^{19,48,79,129} has strengthened the reputation of measuring CNAPs as the most preferred monitoring method of choice.^{14,16,17,19,20,48,99,146} When comparing direct CNAPs with ECOG, direct CNAPs possess higher predictive value of postoperative functionality, with lower false-positive and higher true-positive rates.^{18,146}

Electrocochleography and direct CNAPs have unique disadvantages. Because the recording electrode is placed peripherally, ECOG is unable to provide information about the entirety of the auditory nerve, particularly the more central portions of the auditory pathway.^{18,41,83,98,129} As a result, it is possible to completely transect the nerve centrally without observing any credible change on the ECOG study.^{48,79} Due to its rather invasive nature, ECOG also presents an increased risk of CSF otorrhea due to tympanic membrane perforation during electrode placement.^{49,96,120,149} To circumvent this issue, alternative but viable options include tympanic or extratympanic electrode placement.^{95,113,144} Electrocochleography can prove technically challenging as well. The electrode must be held securely in place; moving it manually or unintentionally can induce changes in the baseline amplitude and latency, thus exacerbating the difficulty of making subsequent comparisons.¹⁴⁹

The disadvantage of using direct CNAPs is mainly practical. In larger tumors, there is very little operating space to place the recording electrode without sacrificing visibility of the surgical field.^{49,54,79,99,120,129,149} As a result, direct CNAPs are generally reserved for patients presenting with smaller tumors.^{42,83}

Conclusions

Implementation of facial and vestibulocochlear nerve IONM, in combination with the development of improved modern microneurosurgical techniques, has led to a dramatic reduction in the morbidity once associated with acoustic neuroma surgery. The facial nerve, in particular, has shown higher rates of preservation with the use of EMGs. The vestibulocochlear nerve, on the other hand, may be important to investigate as an avenue for further improvement. Despite the combined techniques of BAEPs, ECOG, and direct CNAPs, auditory preservation rates do not yet approximate those of facial nerve preservation. Further efforts and investigations are needed to study and incorporate other adjunctive IONM techniques in an attempt to improve preservation of auditory function.

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References

1. Abramson M, Stein BM, Pedley TA, Emerson RG, Wazen JJ: Intraoperative BAER monitoring and hearing preservation in the treatment of acoustic neuromas. **Laryngoscope** **95**:1318–1322, 1985
2. Agnew WF, McCreery DB, Yuen TG, Bullara LA: Histologic and physiologic evaluation of electrically stimulated peripheral nerve: considerations for the selection of parameters. **Ann Biomed Eng** **17**:39–60, 1989
3. Anderson DE, Leonetti J, Wind JJ, Cribari D, Fahey K: Resection of large vestibular schwannomas: facial nerve preservation in the context of surgical approach and patient-assessed outcome. **J Neurosurg** **102**:643–649, 2005
4. Arriaga MA, Chen DA: Facial function in hearing preservation acoustic neuroma surgery. **Arch Otolaryngol Head Neck Surg** **127**:543–546, 2001
5. Arriaga MA, Chen DA, Fukushima T: Individualizing hearing preservation in acoustic neuroma surgery. **Laryngoscope** **107**:1043–1047, 1997
6. Arriaga MA, Luxford WM, Berliner KI: Facial nerve function following middle fossa and translabyrinthine acoustic tumor surgery: a comparison. **Am J Otol** **15**:620–624, 1994
7. Arts HA, Telian SA, El-Kashlan H, Thompson BG: Hearing preservation and facial nerve outcomes in vestibular schwannoma surgery: results using the middle cranial fossa approach. **Otol Neurotol** **27**:234–241, 2006
8. Battista RA, Wiet RJ, Pauwe L: Evaluation of three intraoperative auditory monitoring techniques in acoustic neuroma surgery. **Am J Otol** **21**:244–248, 2000
9. Benecke JE Jr, Calder HB, Chadwick G: Facial nerve monitoring during acoustic neuroma removal. **Laryngoscope** **97**:697–700, 1987
10. Brackmann DE, Owens RM, Friedman RA, Hitselberger WE, De la Cruz A, House JW, et al: Prognostic factors for hearing preservation in vestibular schwannoma surgery. **Am J Otol** **21**:417–424, 2000
11. Briggs RJ, Luxford WM, Atkins JS Jr, Hitselberger WE: Translabyrinthine removal of large acoustic neuromas. **Neurosurgery** **34**:785–791, 1994
12. Cerullo LJ, Grutsch JF, Heiferman K, Osterdock R: The preservation of hearing and facial nerve function in a consecutive series of unilateral vestibular nerve schwannoma surgical patients (acoustic neuroma). **Surg Neurol** **39**:485–493, 1993
13. Ciric I, Zhao JC, Rosenblatt S, Wiet R, O'Shaughnessy B: Suboccipital retrosigmoid approach for removal of vestibular schwannomas: facial nerve function and hearing preservation. **Neurosurgery** **56**:560–570, 2005
14. Colletti V, Bricolo A, Fiorino FG, Bruni L: Changes in directly recorded cochlear nerve compound action potentials during acoustic tumor surgery. **Skull Base Surg** **4**:1–9, 1994
15. Colletti V, Fiorino F: Middle fossa versus retrosigmoid-transmeatal approach in vestibular schwannoma surgery: a prospective study. **Otol Neurotol** **24**:927–934, 2003
16. Colletti V, Fiorino FG, Carner M, Cumer G, Giarbini N, Sacchetto L: Intraoperative monitoring for hearing preservation and restoration in acoustic neuroma surgery. **Skull Base Surg** **10**:187–195, 2000
17. Colletti V, Fiorino FG, Carner M, Tonoli G: Mechanisms of auditory impairment during acoustic neuroma surgery. **Otolaryngol Head Neck Surg** **117**:596–605, 1997
18. Colletti V, Fiorino FG, Mocella S, Policante Z: ECochG, CNAP and ABR monitoring during vestibular Schwannoma surgery. **Audiology** **37**:27–37, 1998
19. Colletti V, Fiorino FG, Sacchetto L: Iatrogenic impairment of hearing during surgery for acoustic neuroma. **Skull Base Surg** **6**:153–161, 1996
20. Danner C, Mastrodimos B, Cueva RA: A comparison of direct eighth nerve monitoring and auditory brainstem response in hearing preservation surgery for vestibular schwannoma. **Otol Neurotol** **25**:826–832, 2004
21. De Seta E, Bertoli G, De Seta D, Covelli E, Filippo R: New development in intraoperative video monitoring of facial nerve: a pilot study. **Otol Neurotol** **31**:1498–1502, 2010
22. Delgado TE, Bucheit WA, Rosenholtz HR, Chrissian S: Intraoperative monitoring of facialis muscle evoked responses obtained by intracranial stimulation of the facialis nerve: a more accurate technique for facialis nerve dissection. **Neurosurgery** **4**:418–421, 1979
23. Dornhoffer JL, Helms J, Hoehmann DH: Hearing preservation in acoustic tumor surgery: results and prognostic factors. **Laryngoscope** **105**:184–187, 1995
24. Dubois MY, Sato S, Chassy J, Macnamara TE: Effects of enflurane on brainstem auditory evoked responses in humans. **Anesth Analg** **61**:898–902, 1982
25. Ebersold MJ, Harner SG, Beatty CW, Harper CM Jr, Quast LM: Current results of the retrosigmoid approach to acoustic neurinoma. **J Neurosurg** **76**:901–909, 1992
26. Eisele DW, Wang SJ, Orloff LA: Electrophysiologic facial nerve monitoring during parotidectomy. **Head Neck** **32**:399–405, 2010
27. Fenton JE, Chin RY, Shirazi A, Fagan PA: Prediction of postoperative facial nerve function in acoustic neuroma surgery. **Clin Otolaryngol Allied Sci** **24**:483–486, 1999
28. Filippo R, De Seta E, Bertoli GA: Intraoperative videomonitoring of the facial nerve. **Am J Otol** **21**:119–122, 2000
29. Filippo R, Pichi B, Bertoli GA, De Seta E: Video-based system for intraoperative facial nerve monitoring: comparison with electromyography. **Otol Neurotol** **23**:594–597, 2002
30. Fischer G, Fischer C, Rémond J: Hearing preservation in acoustic neurinoma surgery. **J Neurosurg** **76**:910–917, 1992
31. Flickinger JC, Kondziolka D, Nirranjan A, Lunsford LD: Results of acoustic neuroma radiosurgery: an analysis of 5 years' experience using current methods. **J Neurosurg** **94**:1–6, 2001
32. Fong B, Barkhoudarian G, Pezeshkian P, Parsa AT, Gopen Q, Yang I: The molecular biology and novel treatments of vestibular schwannomas. A review. **J Neurosurg** **115**:906–914, 2011
33. Gal TJ, Shinn J, Huang B: Current epidemiology and management trends in acoustic neuroma. **Otolaryngol Head Neck Surg** **142**:677–681, 2010
34. Glasscock ME III, Hays JW, Minor LB, Haynes DS, Carrasco VN: Preservation of hearing in surgery for acoustic neuromas. **J Neurosurg** **78**:864–870, 1993
35. Goldbrunner RH, Schlake HP, Milewski C, Tonn JC, Helms J, Roosen K: Quantitative parameters of intraoperative elec-

- tromyography predict facial nerve outcomes for vestibular schwannoma surgery. **Neurosurgery** 46:1140–1148, 2000
36. Gormley WB, Sekhar LN, Wright DC, Kamerer D, Schessel D: Acoustic neuromas: results of current surgical management. **Neurosurgery** 41:50–60, 1997
 37. Grayeli AB, Guindi S, Kalamarides M, El Garem H, Smail M, Rey A, et al: Four-channel electromyography of the facial nerve in vestibular schwannoma surgery: sensitivity and prognostic value for short-term facial function outcome. **Otol Neurotol** 26:114–120, 2005
 38. Grayeli AB, Kalamarides M, Fraysse B, Deguine O, Favre G, Martin C, et al: Comparison between intraoperative observations and electromyographic monitoring data for facial nerve outcome after vestibular schwannoma surgery. **Acta Otolaryngol** 125:1069–1074, 2005
 39. Harner SG, Daube JR, Beatty CW, Ebersold MJ: Intraoperative monitoring of the facial nerve. **Laryngoscope** 98:209–212, 1988
 40. Harner SG, Daube JR, Ebersold MJ: Electrophysiologic monitoring of facial nerve during temporal bone surgery. **Laryngoscope** 96:65–69, 1986
 41. Harper CM: Intraoperative cranial nerve monitoring. **Muscle Nerve** 29:339–351, 2004
 42. Harper CM, Daube JR: Facial nerve electromyography and other cranial nerve monitoring. **J Clin Neurophysiol** 15:206–216, 1998
 43. House JW, Brackmann DE: Facial nerve grading system. **Otolaryngol Head Neck Surg** 93:146–147, 1985
 44. Hughes GB, Bottomy MB, Dickins JR, Jackson CG, Sismanis A, Glasscock ME III: A comparative study of neuropathologic changes following pulsed and direct current stimulation of the mouse sciatic nerve. **Am J Otolaryngol** 1:378–384, 1980
 45. Irving RM, Jackler RK, Pitts LH: Hearing preservation in patients undergoing vestibular schwannoma surgery: comparison of middle fossa and retrosigmoid approaches. **J Neurosurg** 88:840–845, 1998
 46. Isaacson B, Kileny PR, El-Kashlan H, Gadre AK: Intraoperative monitoring and facial nerve outcomes after vestibular schwannoma resection. **Otol Neurotol** 24:812–817, 2003
 47. Isaacson B, Kileny PR, El-Kashlan HK: Prediction of long-term facial nerve outcomes with intraoperative nerve monitoring. **Otol Neurotol** 26:270–273, 2005
 48. Jackson LE, Roberson JB Jr: Acoustic neuroma surgery: use of cochlear nerve action potential monitoring for hearing preservation. **Am J Otol** 21:249–259, 2000
 49. Jaisinghani VJ, Levine SC, Nussbaum E, Haines S, Lindgren B: Hearing preservation after acoustic neuroma surgery. **Skull Base Surg** 10:141–147, 2000
 50. James ML, Husain AM: Brainstem auditory evoked potential monitoring: when is change in wave V significant? **Neurology** 65:1551–1555, 2005
 51. Jannetta PJ, Møller AR, Møller MB: Technique of hearing preservation in small acoustic neuromas. **Ann Surg** 200:513–523, 1984
 52. Jenkins HA: Hearing preservation in acoustic neuroma surgery. **Laryngoscope** 102:125–128, 1992
 53. Karpinos M, Teh BS, Zeck O, Carpenter LS, Phan C, Mai WY, et al: Treatment of acoustic neuroma: stereotactic radiosurgery vs. microsurgery. **Int J Radiat Oncol Biol Phys** 54:1410–1421, 2002
 54. Kartush JM, Larouere MJ, Graham MD, Bouchard KR, Audet BV: Intraoperative cranial nerve monitoring during posterior skull base surgery. **Skull Base Surg** 1:85–92, 1991
 55. Kartush JM, Niparko JK, Bledsoe SC, Graham MD, Kemink JL: Intraoperative facial nerve monitoring: a comparison of stimulating electrodes. **Laryngoscope** 95:1536–1540, 1985
 56. King TT, Morrison AW: Translabyrinthine and transtentorial removal of acoustic nerve tumors. Results in 150 cases. **J Neurosurg** 52:210–216, 1980
 57. Kondziolka D, Lunsford LD, McLaughlin MR, Flickinger JC: Long-term outcomes after radiosurgery for acoustic neuromas. **N Engl J Med** 339:1426–1433, 1998
 58. Kumon Y, Sakaki S, Kohno K, Ohta S, Nakagawa K, Ohue S, et al: Selection of surgical approaches for small acoustic neurinomas. **Surg Neurol** 53:52–60, 2000
 59. Kveton JF: The efficacy of brainstem auditory evoked potentials in acoustic tumor surgery. **Laryngoscope** 100:1171–1173, 1990
 60. Lanman TH, Brackmann DE, Hitselberger WE, Subin B: Report of 190 consecutive cases of large acoustic tumors (vestibular schwannoma) removed via the translabyrinthine approach. **J Neurosurg** 90:617–623, 1999
 61. Lassaletta L, Fontes L, Melcon E, Sarria MJ, Gavilan J: Hearing preservation with the retrosigmoid approach for vestibular schwannoma: myth or reality? **Otolaryngol Head Neck Surg** 129:397–401, 2003
 62. Lee SH, Willcox TO, Buchheit WA: Current results of the surgical management of acoustic neuroma. **Skull Base** 12:189–195, 2002
 63. Legatt AD: Mechanisms of intraoperative brainstem auditory evoked potential changes. **J Clin Neurophysiol** 19:396–408, 2002
 64. Lüders H: Surgical monitoring with auditory evoked potentials. **J Clin Neurophysiol** 5:261–285, 1988
 65. Magnan J, Barbieri M, Mora R, Murphy S, Meller R, Bruzzo M, et al: Retrosigmoid approach for small and medium-sized acoustic neuromas. **Otol Neurotol** 23:141–145, 2002
 66. Mandpe AH, Mikulec A, Jackler RK, Pitts LH, Yingling CD: Comparison of response amplitude versus stimulation threshold in predicting early postoperative facial nerve function after acoustic neuroma resection. **Am J Otol** 19:112–117, 1998
 67. Mangham CA Jr: Retrosigmoid versus middle fossa surgery for small vestibular schwannomas. **Laryngoscope** 114:1455–1461, 2004
 68. Mann WJ, Maurer J, Marangos N: Neural conservation in skull base surgery. **Otolaryngol Clin North Am** 35:411–424, ix, 2002
 69. Manninen PH, Lam AM, Nicholas JF: The effects of isoflurane and isoflurane-nitrous oxide anesthesia on brainstem auditory evoked potentials in humans. **Anesth Analg** 64:43–47, 1985
 70. Marin P, Pouliot D, Fradet G: Facial nerve outcome with a preoperative stimulation threshold under 0.05 mA. **Laryngoscope** 121:2295–2298, 2011
 71. Markand ON: Brainstem auditory evoked potentials. **J Clin Neurophysiol** 11:319–342, 1994
 72. Martin HC, Sethi J, Lang D, Neil-Dwyer G, Lutman ME, Yardley L: Patient-assessed outcomes after excision of acoustic neuroma: postoperative symptoms and quality of life. **J Neurosurg** 94:211–216, 2001
 73. Matthies C, Samii M: Direct brainstem recording of auditory evoked potentials during vestibular schwannoma resection: nuclear BAEP recording. Technical note and preliminary results. **J Neurosurg** 86:1057–1062, 1997
 74. Matthies C, Samii M: Management of vestibular schwannomas (acoustic neuromas): the value of neurophysiology for evaluation and prediction of auditory function in 420 cases. **Neurosurgery** 40:919–930, 1997
 75. Maw AR, Coakham HB, Ayoub O, Butler SR: Hearing preservation and facial nerve function in vestibular schwannoma surgery. **Clin Otolaryngol Allied Sci** 28:252–256, 2003
 76. McCreery DB, Agnew WF, Yuen TGH, Bullara LA: Relationship between stimulus amplitude, stimulus frequency and neural damage during electrical stimulation of sciatic nerve of cat. **Med Biol Eng Comput** 33 (3 Spec No):426–429, 1995
 77. Minahan RE, Mandir AS: Neurophysiologic intraoperative monitoring of trigeminal and facial nerves. **J Clin Neurophysiol** 28:551–565, 2011
 78. Møller AR: Intraoperative neurophysiologic monitoring. **Am J Otol** 16:115–117, 1995

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79. Møller AR: Monitoring auditory function during operations to remove acoustic tumors. **Am J Otol** **17**:452–460, 1996
80. Møller AR, Jannetta PJ: Auditory evoked potentials recorded from the cochlear nucleus and its vicinity in man. **J Neurosurg** **59**:1013–1018, 1983
81. Møller AR, Jannetta PJ: Compound action potentials recorded intracranially from the auditory nerve in man. **Exp Neurol** **74**:862–874, 1981
82. Møller AR, Jannetta PJ: Preservation of facial function during removal of acoustic neuromas. Use of monopolar constant-voltage stimulation and EMG. **J Neurosurg** **61**:757–760, 1984
83. Møller AR, Jho HD, Jannetta PJ: Preservation of hearing in operations on acoustic tumors: an alternative to recording brain stem auditory evoked potentials. **Neurosurgery** **34**:688–693, 1994
84. Mom T, Telischi FF, Martin GK, Stagner BB, Lonsbury-Martin BL: Vasospasm of the internal auditory artery: significance in cerebellopontine angle surgery. **Am J Otol** **21**:735–742, 2000
85. Murphy EK: Use of an infrared camera to improve the outcome of facial nerve monitoring. **Am J Electroneurodiagn Technol** **48**:38–47, 2008
86. Myrseth E, Møller P, Pedersen PH, Lund-Johansen M: Vestibular schwannoma: surgery or gamma knife radiosurgery? A prospective, nonrandomized study. **Neurosurgery** **64**:654–663, 2009
87. Myrseth E, Møller P, Pedersen PH, Vassbotn FS, Wentzel-Larsen T, Lund-Johansen M: Vestibular schwannomas: clinical results and quality of life after microsurgery or gamma knife radiosurgery. **Neurosurgery** **56**:927–935, 2005
88. Nadol JB Jr, Chiong CM, Ojemann RG, McKenna MJ, Martuza RL, Montgomery WW, et al: Preservation of hearing and facial nerve function in resection of acoustic neuroma. **Laryngoscope** **102**:1153–1158, 1992
89. Nadol JB Jr, Levine R, Ojemann RG, Martuza RL, Montgomery WW, de Sandoval PK: Preservation of hearing in surgical removal of acoustic neuromas of the internal auditory canal and cerebellar pontine angle. **Laryngoscope** **97**:1287–1294, 1987
90. Neff BA, Ting J, Dickinson SL, Welling DB: Facial nerve monitoring parameters as a predictor of postoperative facial nerve outcomes after vestibular schwannoma resection. **Otol Neurotol** **26**:728–732, 2005
91. Neu M, Strauss C, Romstöck J, Bischoff B, Fahlbusch R: The prognostic value of intraoperative BAEP patterns in acoustic neurinoma surgery. **Clin Neurophysiol** **110**:1935–1941, 1999
92. Nielsen A: Acoustic tumors: with special reference to end-results and sparing of the facial nerve. **Ann Surg** **115**:849–863, 1942
93. Nissen AJ, Sikand A, Curto FS, Welsh JE, Gardi J: Value of intraoperative threshold stimulus in predicting postoperative facial nerve function after acoustic tumor resection. **Am J Otol** **18**:249–251, 1997
94. Noguchi Y, Komatsuzaki A, Nishida H: Cochlear microphonics for hearing preservation in vestibular schwannoma surgery. **Laryngoscope** **109**:1982–1987, 1999
95. Noguchi Y, Nishida H, Komatsuzaki A: A comparison of extratympanic versus transtympanic recordings in electrocochleography. **Audiology** **38**:135–140, 1999
96. Ojemann RG, Levine RA, Montgomery WM, McGaffigan P: Use of intraoperative auditory evoked potentials to preserve hearing in unilateral acoustic neuroma removal. **J Neurosurg** **61**:938–948, 1984
97. Petrova LD: Brainstem auditory evoked potentials. **Am J Electroneurodiagn Technol** **49**:317–332, 2009
98. Phillips DJ, Kobylarz EJ, De Peralta ET, Stieg PE, Selesnick SH: Predictive factors of hearing preservation after surgical resection of small vestibular schwannomas. **Otol Neurotol** **31**:1463–1468, 2010
99. Piccirillo E, Hiraumi H, Hamada M, Russo A, De Stefano A, Sanna M: Intraoperative cochlear nerve monitoring in vestibular schwannoma surgery—does it really affect hearing outcome? **Audiol Neurootol** **13**:58–64, 2008
100. Pollock BE, Driscoll CL, Foote RL, Link MJ, Gorman DA, Bauch CD, et al: Patient outcomes after vestibular schwannoma management: a prospective comparison of microsurgical resection and stereotactic radiosurgery. **Neurosurgery** **59**:77–85, 2006
101. Pollock BE, Lunsford LD, Kondziolka D, Flickinger JC, Bissonette DJ, Kelsey SF, et al: Outcome analysis of acoustic neuroma management: a comparison of microsurgery and stereotactic radiosurgery. **Neurosurgery** **36**:215–229, 1995
102. Polo G, Fischer C, Sindou MP, Marneffe V: Brainstem auditory evoked potential monitoring during microvascular decompression for hemifacial spasm: intraoperative brainstem auditory evoked potential changes and warning values to prevent hearing loss—prospective study in a consecutive series of 84 patients. **Neurosurgery** **54**:97–106, 2004
103. Post KD, Eisenberg MB, Catalano PJ: Hearing preservation in vestibular schwannoma surgery: what factors influence outcome? **J Neurosurg** **83**:191–196, 1995
104. Prell J, Rachinger J, Scheller C, Alfieri A, Strauss C, Rampp S: A real-time monitoring system for the facial nerve. **Neurosurgery** **66**:1064–1073, 2010
105. Prell J, Rampp S, Romstöck J, Fahlbusch R, Strauss C: Train time as a quantitative electromyographic parameter for facial nerve function in patients undergoing surgery for vestibular schwannoma. **J Neurosurg** **106**:826–832, 2007
106. Randall DA, Wester DC, Hunsaker DH: Reliability of disposable intraoperative facial nerve stimulators. **Laryngoscope** **107**:192–199, 1997
107. Régis J, Pellet W, Delsanti C, Dufour H, Roche PH, Thomassin JM, et al: Functional outcome after gamma knife surgery or microsurgery for vestibular schwannomas. **J Neurosurg** **97**:1091–1100, 2002
108. Roberson J, Senne A, Brackmann D, Hitselberger WE, Saunders J: Direct cochlear nerve action potentials as an aid to hearing preservation in middle fossa acoustic neuroma resection. **Am J Otol** **17**:653–657, 1996
109. Roberson JB Jr, Jackson LE, McAuley JR: Acoustic neuroma surgery: absent auditory brainstem response does not contraindicate attempted hearing preservation. **Laryngoscope** **109**:904–910, 1999
110. Roland JT Jr, Fishman AJ, Golfinos JG, Cohen N, Alexiades G, Jackman AH: Cranial nerve preservation in surgery for large acoustic neuromas. **Skull Base** **14**:85–91, 2004
111. Romstöck J, Strauss C, Fahlbusch R: Continuous electromyography monitoring of motor cranial nerves during cerebellopontine angle surgery. **J Neurosurg** **93**:586–593, 2000
112. Rowed DW, Nedzelski JM: Hearing preservation in the removal of intracanalicular acoustic neuromas via the retrosigmoid approach. **J Neurosurg** **86**:456–461, 1997
113. Ruth RA, Lambert PR, Ferraro JA: Electrocochleography: methods and clinical applications. **Am J Otol** **9** (Suppl):1–11, 1988
114. Sabin HI, Bentivoglio P, Symon L, Cheesman AD, Prasher D, Momma F: Intra-operative electrocochleography to monitor cochlear potentials during acoustic neuroma excision. **Acta Neurochir (Wien)** **85**:110–116, 1987
115. Samii M, Gerganov V, Samii A: Improved preservation of hearing and facial nerve function in vestibular schwannoma surgery via the retrosigmoid approach in a series of 200 patients. **J Neurosurg** **105**:527–535, 2006
116. Samii M, Matthies C: Management of 1000 vestibular schwannomas (acoustic neuromas): hearing function in 1000 tumor resections. **Neurosurgery** **40**:248–262, 1997
117. Sampath P, Holliday MJ, Brem H, Niparko JK, Long DM: Facial nerve injury in acoustic neuroma (vestibular schwannoma) surgery: etiology and prevention. **J Neurosurg** **87**:60–66, 1997
118. Sanna M, Zini C, Mazzoni A, Gandolfi A, Pareschi R, Pansani E, et al: Hearing preservation in acoustic neuroma

- surgery. Middle fossa versus suboccipital approach. **Am J Otol** 8:500–506, 1987
119. Sapmaz E, Kaygusuz I, Alpay HC, Akpolat N, Keles E, Karlidag T, et al: Histopathologic and functional effects of facial nerve following electrical stimulation. **Eur Arch Otorhinolaryngol** 267:607–612, 2010
 120. Schlake HP, Milewski C, Goldbrunner RH, Kindgen A, Riemann R, Helms J, et al: Combined intra-operative monitoring of hearing by means of auditory brainstem responses (ABR) and transtympanic electrocochleography (ECoChG) during surgery of intra- and extrameatal acoustic neuromas. **Acta Neurochir (Wien)** 143:985–996, 2001
 121. Schramm J, Mokrusch T, Fahlbusch R, Hochstetter A: Detailed analysis of intraoperative changes monitoring brain stem acoustic evoked potentials. **Neurosurgery** 22:694–702, 1988
 122. Sekiya T, Möller AR: Avulsion rupture of the internal auditory artery during operations in the cerebellopontine angle: a study in monkeys. **Neurosurgery** 21:631–637, 1987
 123. Selesnick SH, Carew JF, Victor JD, Heise CW, Levine J: Predictive value of facial nerve electrophysiologic stimulation thresholds in cerebellopontine-angle surgery. **Laryngoscope** 106:633–638, 1996
 124. Shelton C, Brackmann DE, House WF, Hitselberger WE: Middle fossa acoustic tumor surgery: results in 106 cases. **Laryngoscope** 99:405–408, 1989
 125. Silverstein H, Rosenberg SI, Flanzer J, Seidman MD: Intraoperative facial nerve monitoring in acoustic neuroma surgery. **Am J Otol** 14:524–532, 1993
 126. Silverstein H, Smouha EE, Jones R: Routine intraoperative facial nerve monitoring during otologic surgery. **Am J Otol** 9:269–275, 1988
 127. Silverstein H, White DW: Continuous electrical stimulation as a helpful adjunct during intraoperative facial nerve monitoring. **Skull Base Surg** 1:127–131, 1991
 128. Silverstein H, Willcox TO Jr, Rosenberg SI, Seidman MD: Prediction of facial nerve function following acoustic neuroma resection using intraoperative facial nerve stimulation. **Laryngoscope** 104:539–544, 1994
 129. Simon MV: Neurophysiologic intraoperative monitoring of the vestibulocochlear nerve. **J Clin Neurophysiol** 28:566–581, 2011
 130. Slattery WH III, Brackmann DE, Hitselberger W: Middle fossa approach for hearing preservation with acoustic neuromas. **Am J Otol** 18:596–601, 1997
 131. Staecker H, Nadol JB Jr, Ojeman R, Ronner S, McKenna MJ: Hearing preservation in acoustic neuroma surgery: middle fossa versus retrosigmoid approach. **Am J Otol** 21:399–404, 2000
 132. Stidham KR, Roberson JB Jr: Hearing improvement after middle fossa resection of vestibular schwannoma. **Otol Neurotol** 22: 917–921, 2001
 133. Stockard JJ, Sharbrough FW, Tinker JA: Effects of hypothermia on the human brainstem auditory response. **Ann Neurol** 3: 368–370, 1978
 134. Strauss C: The facial nerve in medial acoustic neuromas. **J Neurosurg** 97:1083–1090, 2002
 135. Strauss C, Bischoff B, Neu M, Berg M, Fahlbusch R, Romstöck J: Vasoactive treatment for hearing preservation in acoustic neuroma surgery. **J Neurosurg** 95:771–777, 2001
 136. Strauss C, Fahlbusch R, Romstöck J, Schramm J, Watanabe E, Taniguchi M, et al: Delayed hearing loss after surgery for acoustic neuromas: clinical and electrophysiological observations. **Neurosurgery** 28:559–565, 1991
 137. Taha JM, Tew JM Jr, Keith RW: Proximal-to-distal facial amplitude ratios as predictors of facial nerve function after acoustic neuroma excision. **J Neurosurg** 83:994–998, 1995
 138. Theodosopoulos PV, Pensak ML: Contemporary management of acoustic neuromas. **Laryngoscope** 121:1133–1137, 2011
 139. Tonn JC, Schlake HP, Goldbrunner R, Milewski C, Helms J, Roosen K: Acoustic neuroma surgery as an interdisciplinary approach: a neurosurgical series of 508 patients. **J Neurol Neurosurg Psychiatry** 69:161–166, 2000
 140. Tufarelli D, Meli A, Alesii A, De Angelis E, Badaracco C, Falcioni M, et al: Quality of life after acoustic neuroma surgery. **Otol Neurotol** 27:403–409, 2006
 141. Watanabe E, Schramm J, Strauss C, Fahlbusch R: Neurophysiologic monitoring in posterior fossa surgery. II. BAEP-waves I and V and preservation of hearing. **Acta Neurochir (Wien)** 98:118–128, 1989
 142. Wazen JJ: Intraoperative monitoring of auditory function: experimental observations and new applications. **Laryngoscope** 104: 446–455, 1994
 143. Wiegand DA, Ojemann RG, Fickel V: Surgical treatment of acoustic neuroma (vestibular schwannoma) in the United States: report from the Acoustic Neuroma Registry. **Laryngoscope** 106:58–66, 1996
 144. Winzenburg SM, Margolis RH, Levine SC, Haines SJ, Fournier EM: Tympanic and transtympanic electrocochleography in acoustic neuroma and vestibular nerve section surgery. **Am J Otol** 14:63–69, 1993
 145. Yamakami I, Oka N, Yamaura A: Intraoperative monitoring of cochlear nerve compound action potential in cerebellopontine angle tumour removal. **J Clin Neurosci** 10:567–570, 2003
 146. Yamakami I, Yoshinori H, Saeki N, Wada M, Oka N: Hearing preservation and intraoperative auditory brainstem response and cochlear nerve compound action potential monitoring in the removal of small acoustic neurinoma via the retrosigmoid approach. **J Neurol Neurosurg Psychiatry** 80:218–227, 2009
 147. Yates PD, Jackler RK, Satar B, Pitts LH, Oghalai JS: Is it worthwhile to attempt hearing preservation in larger acoustic neuromas? **Otol Neurotol** 24:460–464, 2003
 148. Youssef AS, Downes AE: Intraoperative neurophysiological monitoring in vestibular schwannoma surgery: advances and clinical implications. **Neurosurg Focus** 27(4):E9, 2009
 149. Zappia JJ, Wiet RJ, O'Connor CA, Martone L: Intraoperative auditory monitoring in acoustic neuroma surgery. **Otolaryngol Head Neck Surg** 115:98–106, 1996
 150. Zeitouni AG, Hammerschlag PE, Cohen NL: Prognostic significance of intraoperative facial nerve stimulus thresholds. **Am J Otol** 18:494–497, 1997

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